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In search for new players of the oxidative stress network by phenotyping an *Arabidopsis* T-DNA mutant collection on reactive oxygen species-eliciting chemicals

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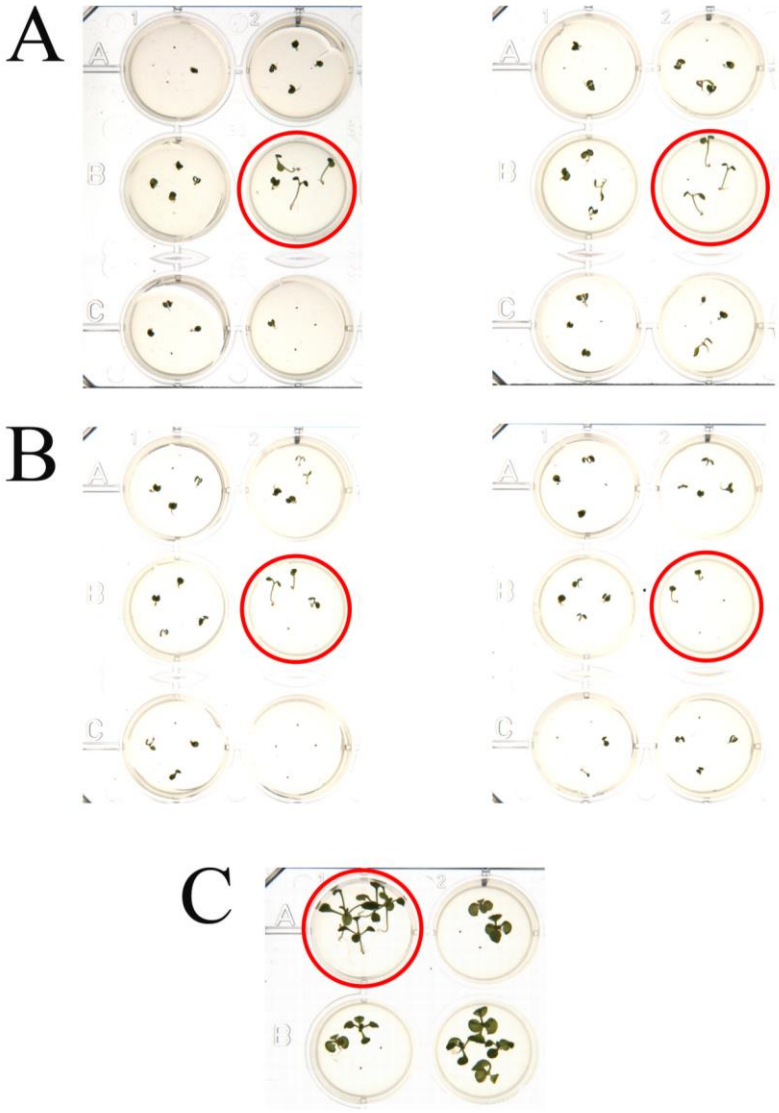
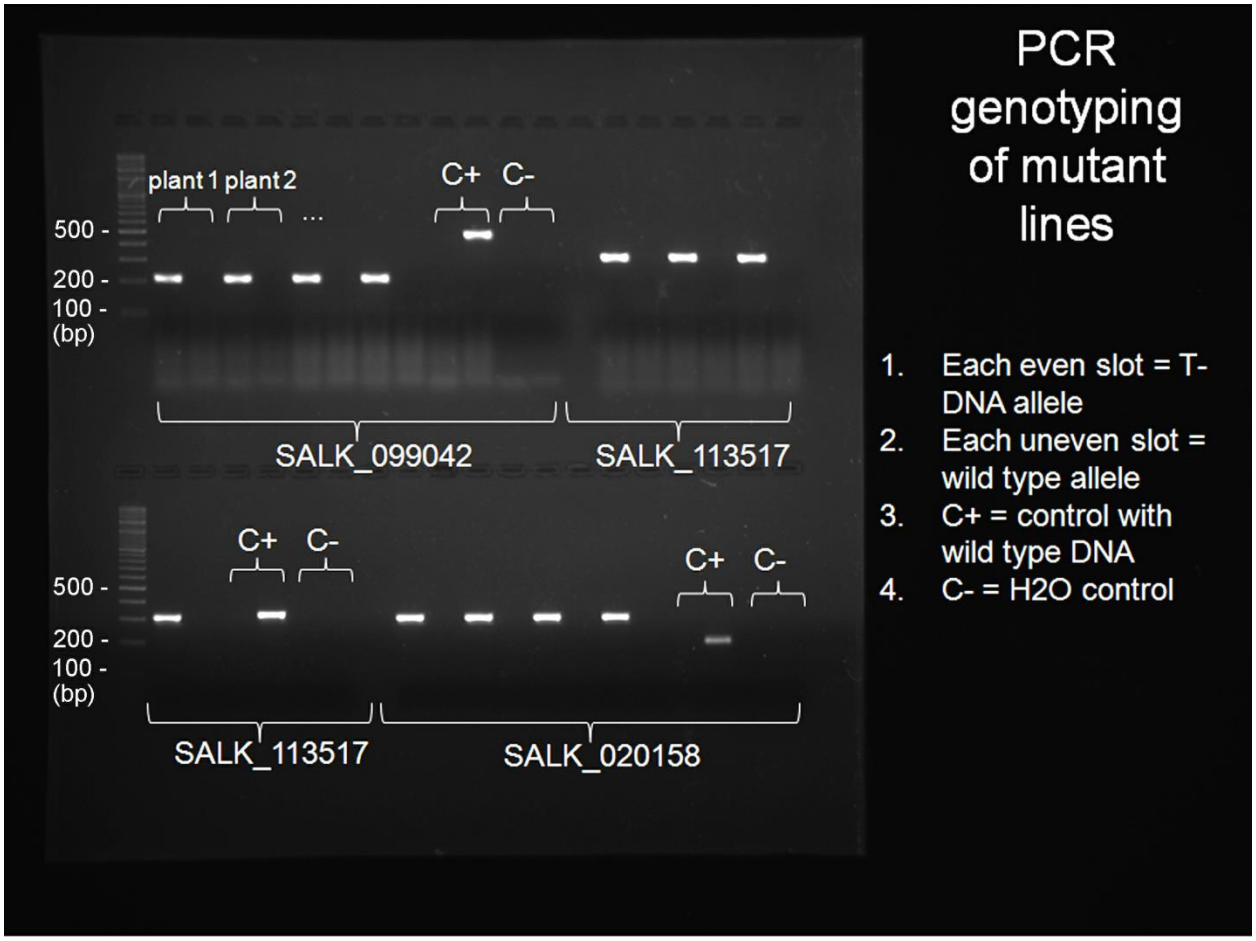


Supplemental Figure 1. Phenotype of the "long hypocotyl" mutant in different culture conditions. Wells with "long hypocotyl" seedlings are encircled in red; the other wells harbor other T-DNA insertion lines. Panel A: Seedlings grown on AT-containing medium. Panel B: Seedlings grown on MV-containing medium. Panel C: Seedlings grown on normal MS medium.



Supplemental Figure 2. PCR genotyping of SALK mutant lines. Four plants per line were analyzed. The first lanes in the upper and lower parts of the gel contain molecular marker DNA (O'gene Ruler, 100bp; Fermentas GmbH, St. Leon-Rot, Germany); every even-numbered slot is loaded with the product of a PCR reaction for T-DNA detection; every odd-numbered slot contains wild-type allele fragments. DNA from Col-0 wild-type plants was used as a positive control, while water was used in negative controls. DNA from each plant was subjected to two PCR reactions, one detecting the T-DNA allele and one the wild-type allele. The reaction products were loaded on adjacent slots for each plant. As seen in the figure, all mutant plants tested were homozygous for the T-DNA insertions (i.e., no amplicons were visible for the wild-type alleles).

primer	sequence
099042F	CAGCATCATCAATAGCTGAGAAAC
099042GR	TTTGGAAGTTCTGGGAGCAG
113517FG	CGTCCTGAAGACACAGATGC
113517R	CCAGTCACAACCTCCCAAC
020158F	GAAGTTCTTCGAGCGTCTCC
020158GR	TTAAACGACCTTTCGGGTTG

TDNA-primer	TGGACCGCTTGCTGCAAC

Supplemental Table 1. The table lists the primers utilized to genotype the different SALK lines. The primers marked with "G" (099042GR, 113517FG and 020158GR) were used to detect wild-type alleles in combination with 099042F, 113517R and 020158F. The T-DNA-primer together with 099042F, 113517R and 020158F, respectively, was used to detect T-DNA insertions.